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Synthesis of a 3-aminopiperidin-2,5-dione as a conformationally constrained surrogate of the Ala-Gly dipeptide

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Abstract—The preparation of the Boc-{Ala-Gly}-OBn pseudopeptide 4 is reported. The key intermediate, aminoester 5b, was obtained by a cross-coupling reaction of alaninezinc iodide 6 and the thioester of glycine 9. © 2000 Elsevier Science Ltd. All rights reserved.

Compounds that possess a 3-amino-2-piperidone nucleus often show important biological activities. For instance, compound 1 (Fig. 1) has been reported as a serine protease inhibitor, and compound 2 is an angiotensin converting enzyme (ACE) inhibitor. Compound 1 consists of the thiazolopiperidone bicyclic system known to be a β -turn mimetic, 3 and cyclo-arginine. Compound 3, a thrombin inhibitor, 4 is a combination of compound 2 and cyclo-arginine. We present here the preparation of 3-aminopiperidin-2,5-dione 4. Compound 4 is a new conformationally constrained Ala-Gly surrogate, functionalised at C5, in which the conformational restriction is caused by the cyclisation between the $C\alpha$ of alanine and the nitrogen atom of glycine. We also intend to use diketopiperidine 4 for the preparation of other

ACE inhibitor Serine protease inhibitor CO₂Bn 3 Thrombine inhibitor Boc-{Ala-Gly}-OBn

pseudodipeptides by using the reactivity of the C5 carbonyl

We considered that 2,5-dioxopiperidine 4 could be prepared by lactamisation of an appropriate aminoester, such as 5 (Fig. 2). In turn, compound 5 could be obtained from simple aminoacid precursors, by acylation of an alanine β-anion equivalent.

Such acylations are usually best achieved by palladiumcatalysed coupling of the organozinc derivative 6a with acid chlorides in benzene/dimethylacetamide as solvent.⁵ Although we have successfully coupled the Fmoc-protected glycine acid chloride with the iodoalanine-derived zinc reagent 6a, to give ketone 5a (Scheme 1), the yield was very poor (26%).6

Therefore, we decided to apply the recently reported palladium-catalysed acylation of simple organozinc iodides with thioesters, which proceeds well in THF, in contrast with acylation using acid chlorides which is generally inefficient in this solvent. In order to assess the reactivity towards thioesters of zinc reagent 6b, obtained from protected iodoalanine 7 by zinc insertion,8 we first treated compound 6b with ethyl acetylthiolate.9 The coupling,

Figure 2.

Keywords: aminolactam; diketopiperidine; dioxopiperidine; peptide mimetic; β-turn mimetic.

Figure 1.

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Scheme 1.

Scheme 2.

using $PdCl_2(PPh_3)_2$ as the catalyst, gave the expected ketoester **8** in 39% yield. The modest yield partly reflects the use of the less reactive ethyl thioester.

Once the reactivity of **6b** towards thioesters had been established, we studied its coupling to the thioester derived from Cbz-glycine **9**. The reaction gave the expected ketoester **5b** in 39% yield, together with the protonated by-product Boc-Ala-OMe (51%). In order to improve this result, we explored various reaction conditions, and identified a catalyst prepared in situ from $Pd_2(dba)_3$ and $P(o\text{-tolyl})_3$, as optimal (Scheme 2).

Once the preparation of compound **5b** had been optimised, its lactamisation to obtain compound **11** was first attempted by hydrogenolysis of the Cbz group. The only compound isolated from the reaction was identified as pyrazine **10**, and resulted from the dimerisation and aromatisation of the deprotected aminoketone (Scheme 3).

Since all attempts to prepare ethylene acetal **12** were unsuccessful ¹¹ (Scheme 4), we treated compound **5** with L-Selectride, which yielded lactone *trans-***13**.^{6,12} The most characteristic ¹H NMR signals of lactone *trans-***13** were a broad doublet at δ 4.22 and a broad singlet at δ 4.73, corresponding to H-3 and H-5, respectively. The *trans* stereochemistry was assigned on the basis of the small chemical shift difference between the protons at C-4 ($\Delta \delta \approx 0.16$).⁶ Hydrogenolysis of the Cbz group of lactone **13** gave the desired 5-hydroxylactam **14** in 15% yield, which was identified by comparison of its spectral data to that reported.¹³

As an alternative to promote lactamisation over dimerisation, we activated the methyl ester **5b** by transforming it into a pentafluorophenyl ester, via acid 15 (Scheme 5). To our satisfaction, hydrogenolysis of the Cbz group of the pentafluorophenyl derivative 16 gave 3-amino-2,5-dioxopiperidine 11 in 79% yield. The most relevant ¹³C NMR data of lactam 11 were the C3 methine carbon at δ 47.8, and two methylene signals at δ 42.3 and 51.5, corresponding to C4 and C6, in addition to the new lactam carbonyl signal at δ 170.9. Finally, alkylation of lactam 11 with benzyl bromoacetate using LHMDS as the base yielded the target piperidin-2,5-dione 4, albeit in poor yield. In the ¹H NMR spectrum, compound 4 showed a characteristic AB system at δ 3.78 and 4.25 (J=17 Hz), corresponding to the exocyclic NCH₂ protons (α -H), and two singlets at δ 4.25 and 5.12 corresponding to the 6-H and benzyl protons, respectively. In the 13 C NMR spectrum C6 and C α were coincident at δ 48.4.

With the synthesis of compound 4, we have demonstrated the usefulness of organozinc chemistry for the preparation of 3-amino-2,5-dioxopiperidines. In future work, we intend to study some applications of compound 4, both as a constrained surrogate of the Ala-Gly dipeptide and as a precursor of other pseudopeptides.

1. Experimental

1.1. General

Melting points were determined in a capillary tube on a

$$\begin{array}{c} \delta_{H}=8.32\text{ s}\\ \delta_{C}=150.8 \end{array}$$
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$$\begin{array}{c} \delta_{H}=8.32\text{ s}\\ \delta_{C}=150.8 \end{array}$$

Scheme 4.

Scheme 5.

Büchi apparatus. Optical rotations were measured with a Perkin-Elmer 241 polarimeter, at 23°C. IR spectra were recorded on a Nicolet FT-IR spectrophotometer. ¹H and ¹³C NMR spectra were recorded in CDCl₃ unless otherwise indicated, on a Varian Gemini-300 instrument. Chemical shifts are expressed in parts per million (δ) relative to Me₄Si. Mass spectra were determined on a Hewlett-Packard 5988A mass spectrometer by electronic impact (EIMS). TLC was performed on SiO₂ (silica gel 60 F254, Macherey-Nagel) and developed with the eluent described for column chromatography. The spots were located with ninhydrin, potassium hexachloroplatinate, anisaldehyde, or KMnO₄. Purification of reagents and solvents was performed according to standard methods. Microanalyses were performed on a Carlo Erba 1106 analyzer at the Serveis Científico-Tècnics (Universitat de Barcelona). Protected iodoalanine 7 was prepared by the literature method.8

1.1.1. Methyl (S)-2-(tert-butoxycarbonylamino)-4-oxopentanoate (8). To a suspension of Zn (1.17 g, 18 mmol) in dry THF (2 ml) 1,2-dibromoethane (77 μ l, 0,9 mmol) was added and the mixture was stirred at 45°C under N₂ atmosphere for 20 min. The mixture was cooled, TMSCl (22 μ l, 0.18 mmol) was added, and the suspension was stirred for 20 min at rt. A solution of protected iodoalanine 7 (987 mg, 3 mmol) in dry THF (2 ml) was added via syringe. The mixture was stirred at 45°C until complete consumption of the substrate (tlc control, 1.45 h). S-Ethyl-

sulfanylacetic acid (470 μl, 4.5 mmol) and PdCl₂(PPh₃)₂ (111 mg, 0.15 mmol) were sequentially added to the organozinc derivative thus obtained. After stirring at 45°C for 4.5 h, the reaction was quenched by addition of AcOEt and 1 M NH₄Cl. The mixture was filtered through Celite to remove the excess of Zn, the solution was diluted with AcOEt, and washed with brine. The organic extracts were dried and evaporated to give a yellow oil which was chromatographed (hexane: AcOEt, 7:3) to obtain the desired ketoester 8 (lower R_f , 279 mg, 39%) and a small proportion of Boc-Ala-OMe (higher R_f , 43 mg, 7%). Ketoester 8: $[\alpha]_D^{22} = +32.7$ (c 1, CHCl₃). IR (NaCl) 1718 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.48 (s, 9H, C(CH₃)₃), 2.17 (s, 3H, H-5), 2.96 (dd, J=18, 4 Hz, 1H, H_A-3), 3.18 (dd, J=18, 4 Hz, 1H, H_B-3), 3.73 (s, 3H, CO₂CH₃), 4.49 (t, *J*=4 Hz, 1H, H-2), 5.51 (d, J=8 Hz, 1H, NH). ¹³C NMR (CDCl₃) 28.3 (C(CH₃)₃), 29.9 (C-5), 45.4 (C-3), 49.4 (C-2), 52.6 (CO_2CH_3) , 80.0 $(C(CH_3)_3)$, 155.5 (NH-CO-Boc), 171.8 (CO_2CH_3) , 206.6 (C-4). EIMS m/z (%) 245 (M⁺, 0.1), 130 (12), 86 (28), 57 (100). Anal. Calcd for C₁₁H₁₉NO₅: C, 53.87; H, 7.81; N, 5.71. Found: C, 53.40; H, 7.93; N, 5.76.

1.1.2. S-Phenyl N-benzyloxycarbonylaminoethanothioate (9). To a solution of Z-Gly (20 g, 95.6 mmol) in dry THF (500 ml) cooled at -10° C and under N_2 atmosphere, DMAP (2.92 g, 23.9 mmol) was added. After 10 min, DCC (49.2 g, 239 mmol) was added, and the formation of a white precipitate was observed. After 30 min at -10° C, recently

1.1.3. Methyl (S)-5-(benzyloxycarbonylamino)-2-(tertbutoxycarbonylamino)-4-oxopentanoate (5b). Operating as for the preparation of 8, from Zn (588.5 mg, 9 mmol) activated with 1,2-dibromoethane (38 µl, 0.45 mmol) and TMSCl (11 µl, 0.09 mmol), iodoalanine 7 (493 mg, 1.5 mmol), dry THF (2 ml), thioester 9 (677 mg, 2.25 mmol), $Pd_2(dba)_3$ (34 mg, 0.03 mmol), and $P(o-tol)_3$ (45.65 mg, 0.15 mmol), a yellow oil was obtained, which was chromatographed (hexane:AcOEt, 8:2 and 2:8) to obtain the desired aminoester 5b (288 mg, 49%), Boc-Ala-OMe (92 mg, 30%), and the unaltered excess thioester 9 (305 mg). **Aminoester 5b**: $[\alpha]_D^{22} = +22.4$ (c 1, CHCl₃). Mp 80-81°C (AcOEt). IR (NaCl) 1760, 1750 and 1725 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.43 (s, 9H, C(CH₃)₃), 2.95 (dd, J=17, 4 Hz, 1H, H_A-3), 3.09 (br d, J=17 Hz, 1H, H_B-3), 3.72 (s, 3H, CO_2CH_3), 4.11 (d, J=5 Hz, 2H, H-5), 4.56 (br s, 1H, H-2), 5.24 (s, 2H, $CH_2C_6H_5$), 5.48 (d, J=6 Hz, 2H, NH), 7.27 (s, 5H, C₆H₅); ¹³C NMR (CDCl₃) 28.3 (C(CH₃)₃), 41.9 (C-3), 49.4 (C-2), 50.6 (C-5), 52.8 (CO₂CH₃), 67.1 (CH₂C₆H₅), 80.3 (C(CH₃)₃), 128.1 (C₆H₅o and -p), 128.4 (C₆H₅-m), 136.2 (C₆H₅-ipso), 155.4 (NH-CO-Boc), 156.1 (NH–CO-Cbz), 171.5 (CO₂CH₃), 203.6 (C-4). EIMS m/z (%) 395 (M⁺+1, 0.1), 294 (3), 174 (14), 146 (14), 91 (100), 57 (63). Anal. Calcd for C₁₉H₂₆N₂O₇: C, 57.86; H, 6.64; N, 7.10. Found: C, 57.70; H, 6.60; N, 6.76.

1.1.4. (S,S)-2,5-Bis(2-tert-butoxycarbonylamino-2-methoxycarbonylethyl)pyrazine (10). To a solution of ketoester **5b** (100 mg, 0.25 mmol) in CH₃OH (1.3 ml) a catalytic amount of 10% Pd/C was added and the dispersion was hydrogenated in a shaker at rt for 24 h. The reaction mixture was filtered through Celite®, the solvent was evaporated, and the residue was chromatographed (hexane:AcOEt, 3:7) to obtain pyrazine 10 (75 mg, 25%) as a yellow oil. $[\alpha]_D^{22}$ = +27.3 (c 1, CHCl₃). IR (NaCl) 1748, 1716 and 1680 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.42 (s, 9H, C(CH₃)₃), 3.29 (d, J=5 Hz, 2H, H-3'), 3.71 (s, 3H, CO₂CH₃), 4.68-4.74 (m,1H, H-2'), 5.55 (d, J=8 Hz, 1H, NH), 8.32 (s, 1H, H-3, and H-6 Ar); ¹³C NMR (CDCl₃) 28.2 (C(CH₃)₃), 36.5 (C-1'), 52.4 (CO₂CH₃), 52.6 (C-2'), 80.0 (C(CH₃)₃), 143.9 (C-3 and C-6), 150.8 (C-2 and C-5), 155.2 (NH-CO-Cbz), 171.9 (CO_2CH_3). EIMS m/z (%) 482 (M^+ , 0.1), 426 (3), 353 (7), 57 (100).

1.1.5. (3*S*,5*R*)-5(*N*-Benzyloxycarbonylaminomethyl)-3-(*tert*-butoxycarbonylamino)-δ-butyrolactone (13). To a

solution of ketoester 5 (125 mg, 0.31 mmol) in dry THF (1.6 ml) cooled at -78°C and under N_2 atmosphere, L-Selectride (0.37 ml, 0.37 mmol) was slowly added. After stirring for 2 h at -78° C, the reaction was quenched by addition of 1 M NH₄Cl. The THF was evaporated, the residue was dissolved in AcOEt and was washed with 1 M NH₄Cl and with brine. The organic extracts dried and evaporated yielded an oil which was chromatographed (hexane:AcOEt, 2:1) to obtain lactone trans-13 (40 mg, 45%). $[\alpha]_D^{22} = -22.4$ (c 0.5, CHCl₃). Mp 143–145°C (AcOEt). IR (NaCl) 3400 (NH), 1706 and 1802 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.43 (s, 9H, C(CH₃)₃), 2.27–2.37 (m, 1H, H-4a), 2.48 (br t, J=9 Hz, 1H, H-4b), 3.34 (dt, J=15, 6 Hz, 1H, H-1'a), 3.53 (ddd, J=15, 6 and 4 Hz, 1H, H-1'b), 4.22 (br d, J=7.5 Hz, 1H, H-3), 4.73 (br s, 1H, H-5), 5.10 (s, 3H, NH-Boc, $CH_2C_6H_5$), 5.20 (t, J=6 Hz, 1H, NH-Cbz), 7.35 (s, 5H, C_6H_5); ¹³C NMR (CDCl₃) 28.2 (C(CH_3)₃), 31.5 (C-4), 44.7 (C-1'), 49.5 (C-3), 67.1 (CH₂C₆H₅), 77.1 (C-5), 80.8 $(CO_2(CH_3)_3)$, 128.1 (C_6H_5-o) , 128.2 (C_6H_5-p) , 128.5 (C₆H₅-m), 136.0 (C₆H₅-ipso), 155.1 (NH–CO-Boc), 156.5 (NH–CO-Cbz), 174.8 (C-2). EIMS *m/z*(%) 308 $(M^+-CO_2(CH_3)_3, 2), 201 (3), 91 (72), 57 (100).$ Anal. Calcd for a $C_{18}H_{24}N_2O_6$: C, 59.33; H, 6.64; N, 7.69. Found: C, 59.28; H, 6.76; N, 7.48.

1.1.6. (3*S*,5*R*)-3-tert-Butoxycarbonylamino-5-hydroxy-2-piperidone (14). To a solution of lactone 13 (80 mg, 0.21 mmol) in CH₃OH (1 ml) a catalytic amount 10% of Pd/C was added, and the dispersion was hydrogenated at P_{atm} and room temperature for 2 h. The mixture was filtered through Celite[®], the solvent was evaporated, and the resulting oil was chromatographed (AcOEt:CH₃OH, 94:6) to obtain 5-hydroxylactam 14¹³ (10 mg, 19%). ¹H NMR (CDCl₃) 1.46 (s, 9H, C(CH₃)₃), 1.65–1.85 (m, 1H, H-4a), 2.85 (br s, 1H, H-4b), 3.24–3.52 (m, 2H, H-6), 4.05 (br s, 1H, H-3), 4.20 (br s, 1H, H-5), 5.59 (d, J=4 Hz, NH-Boc), 6.62 (br s, NH-lactam); ¹³C NMR (CDCl₃) 28.4 (C(CH₃)₃), 37.0 (C-4), 49.8 (C-6), 51.6 (C-3), 63.6 (C-5), 80.4 (C(CH₃)₃), 155.2 (NH-Boc), 170.9 (C-2). EIMS m/z (%) 175 (M⁺-C(CH₃)₃, 11), 157 (12), 57 (100).

1.1.7. (S)-5-(Benzyloxycarbonylamino)-2-(tert-butoxycarbonylamino)-4-oxopentanoic acid (15). To a solution of ketoester **5** (150 mg, 0.38 mmol) in CH₃OH (0.95 ml) cooled at 0°C, 1 M NaOH (0.38 ml) was slowly added. After 1 h the reaction mixture was acidified by addition of AcOH at 0°C, and the solvent was removed under reduced pressure. The residue was dissolved in AcOEt and was washed with H₂O and brine to give acid 15 as a red foam (144 mg, 98%). IR (NaCl) 3600 (CO₂H), 1790 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.44 (s, 9H, C(CH₃)₃), 2.96 (dd, J=18, 6 Hz, 1H, H_A -3), 3.12 (br d, J=18 Hz, 1H, H_B -3), 4.07–4.17 (m, 2H, H-5), 4.58 (br s, 1H, H-2), 5.11 (s, 2H, CH₂C₆H₅),5.48 (br s, 1H, NH), 5.56 (d, J=4 Hz, 1H, NH), 7.35 (s, 5H, C_6H_5); ¹³C NMR (CDCl₃) 28.3 (C(CH₃)₃), 41.6 (C-3), 49.3 (C-2), 50.4 (C-5), 67.2 (CH₂C₆H₅), 80.5 (C(CH₃)₃), 128.1 $(C_6H_5-o \text{ and } m)$, 128.4 (C_6H_5-p) , 136.0 (C_6H_5-ipso) , 155.6 (NH-CO-Boc), 156.4 (NH-CO-Cbz), 174.4 (CO₂H), 204.5 (C-4). EIMS m/z (%) 381 (M⁺+1, 0.1), 91 (100), 57 (47).

1.1.8. Pentafluorophenyl (S)-5-benzyloxycarbonylamino-2-tert-butoxycarbonylamino-4-oxopentan-oate (16). To a solution of acid 15 (650 mg, 1.71 mmol) in dry THF

(8.55 ml) cooled at 0°C and under N₂ atmosphere, DCC (352 mg, 1.71 mmol) was added. After 10 min pentafluorophenol (346 g, 1.88 mmol) was added, and the temperature was left to rise to rt. After stirring for 5 h, the reaction mixture was filtered and the solvent was evaporated under reduced pressure. The resulting yellow oil was chromatographed (hexane:AcOEt, 7:3) to yield the activated ester 16 (766 mg, 85%). IR (NaCl) 1713 (CO), 1520 (CF) cm⁻¹; ¹H NMR (CDCl₃) 1.45 (s, 9H, C(CH₃)₃), 3.14 (dd, J=18, 4 Hz, 1H, H_A -3), 3.31 (dd, J=18, 4 Hz, 1H, H_B -3), 4.13 (t, J=5 Hz, 2H, H-5), 4.94 (t, J=5 Hz, 1H, H-2), 5.12 (s, 2H, $CH_2C_6H_5$), 5.41 (br s, 1H, NH), 5.55 (d, J=8 Hz, 1H, NH), 7.35 (s, 5H, C_6H_5); ¹³C NMR (CDCl₃) 28.2 (C(CH_3)₃), 41.9 (C-3), 49.0 (C-2), 50.4 (C-5), 67.3 ($CH_2C_6H_5$), 81.0 $(C(CH_3)_3)$, 128.2 $(C_6H_5-o \text{ and } -m)$, 128.5 (C_6H_5-p) , 135.9 (C₆H₅-ipso), 139–143 (C₆F₅), 155.1 (NH–CO-Boc), 156.2 (NH–CO-Cbz), 167.5 ($CO_2C_6F_5$), 203.2 (C-4). MS m/z (%) 527 (M+-F, 0.1), 184 (13), 136 (16), 91 (96), 57 (100).

1.1.9. (S)-3-tert-Butoxycarbonylaminopiperidin-2,5-dione (11). To a solution of pentafluorophenyl ester 16 (650 mg, 1.19 mmol) in CH₃OH (119 ml) a catalytic amount of 10% Pd/C was added, and the mixture was hydrogenated for 1.5 h under atmospheric pressure. The reaction mixture was filtered through Celite®, the solvent was evaporated, and the oily residue was chromatographed (hexane:AcOEt, 1:9) to obtain 5-oxolactam 11 (215 mg, 79%). IR (NaCl) 3300 (NH), 1682 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.46 (s, 9H, $C(CH_3)_3$), 2.53 (dd, J=17, 13 Hz, 1H, H-6a), 3.25 (dd, J=17, 4.5 Hz, 1H, H-6b), 3.85 (dd, J=19, 5 Hz, 1H,H-4a), 4.00 (d, J=19 Hz, 1H, H-4b), 4.50 (br t, J=6 Hz, 1H, H-3), 5.59 (d, J=4.5 Hz, NH–Boc), 6.62 (br s, NH-lactam); ¹³C NMR (CDCl₃) 28.3 (C(*C*H₃)₃), 42.3 (C-4), 47.8 (C-3), 51.5 (C-6), 80.4 (C(CH₃)₃), 155.2 (NH–CO-Boc), 170.9 (CO lactam), 202.3 (C-5). EIMS m/z (%) 155 (M⁺-OC(CH₃)₃, 13), 111 (38), 57 (100). Anal. Calcd for C₁₀H₁₆N₂O₄: C, 52.63; H, 7.01; N, 12.28. Found: C, 53.03; H, 6.98; N, 12.30.

1.1.10. (S)-N-Benzyloxycarbonylethyl-3-(N-tert-butoxycarbonylamino)piperidin-2,5-dione (4). To a solution of lactam 11 (60 mg, 0.26 mmol) in dry THF (1 ml) cooled at -78° C and under N₂ atmosphere, LHMDS (0.26 ml, 0.26 mmol) was added. After stirring for 30 min, benzyl bromoacetate (0.06 ml, 0.39 mmol) was added, and the temperature was left to raise to rt. After 5 h, the reaction was quenched by addition of saturated aqueous NH₄Cl and was partitioned with AcOEt/H₂O. The organic extracts, dried and evaporated gave a yellow oil which was chromatographed (hexane: AcOEt, 7:3) to obtain the target lactam 4 (13 mg, 13%). IR (NaCl) 3500 (NH), 1741, 1728 and 1676 (CO) cm⁻¹; ¹H NMR (CDCl₃) 1.46 (s, 9H, C(CH₃)₃), 2.58 (dd, J=16, 13 Hz, 1H, H-4a), 3.25 (dd, J=16, 4.5 Hz, 1H,H-4b), $3.78(d, J=17 Hz, 1H, NCH_A)$, 4.25(d, J=17 Hz, 1H,NCH_B), 4.25 (s, 2H, H-6), 4.45-4.56 (m, 1H, H-3), 5.12 (s, 2H, CH₂Ph), 5.59 (d, *J*=4 Hz, NH-Boc); ¹³C NMR (CDCl₃) 28.3 (C(CH₃)₃), 42.3 (C-4), 48.4 (C-3 and C-α), 58.0 (C-6),

67.6 (CH₂Ph), 81.3 (C(CH₃)₃), 128.4 (Ph-mand Ph-p), 156.1 (NH-Boc), 170.1 (C-2), 201.3 (C-5). EIMS m/z (%) 375 (M⁺ – 1, 0.1), 303 (1), 259 (28), 91 (95), 57 (100). Anal. Calcd for C₁₉H₂₄N₂O₆: C, 60.63; H, 6.43; N, 7.44. Found: C, 60.52; H, 6.57; N, 7.21.

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References

- Tamura, S. Y.; Goldman, E. A.; Brunck, T. K.; Ripka, W. C.; Semple, J. E. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 331–336.
- Thorsett, E. D.; Harris, E. E.; Aster, S. D.; Peterson, E. R.; Snyder, J. P. *J. Med. Chem.* 1986, 29, 251–260.
- For reviews, see: (a) Nagai, U.; Sato, K.; Nakamura, R.; Kato, R. *Tetrahedron* 1993, 49, 3577–3592. (b) Hanessian, S.; McNaughton-Smith, G.; Lombart, H.-G.; Lubell, W. D. *Tetrahedron* 1997, 53, 12789–12854.
- Semple, J. E.; Rowley, D. C.; Brunck, T. K.; Ha-Uong, T.; Minami, N. K.; Owens, T. D.; Tamura, S. Y.; Goldman, E. A.; Siev, D. V.; Ardecky, R. J.; Carpenter, S. H.; Ge, Y.; Richard, B. M.; Nolan, T. G.; Hakanson, K.; Tulinsky, A.; Nutt, R.; Ripka, W. C. J. Med. Chem. 1996, 39, 4531–4536.
- Jackson, R. F. W. Preparation and Use of Organozinc Halides. In *Organozinc Reagents: A Practical Approach*; Knochel, P., Jones, P., Eds.; Oxford University Press: Oxford, 1999; pp 37–56
- Jackson, R. F. W.; Rettie, A. B.; Wood, A.; Wythes, M. J. J. Chem. Soc., Perkin Trans. 1 1994, 1719–1726.
- 7. Tokuyama, H.; Yokoshima, S.; Yamashita, T.; Fukuyama, T. *Tetrahedron Lett.* **1998**, *39*, 3189–3192.
- 8. Bajgrowicz, J. A.; El Hallaoui, A.; Jacquier, R.; Pigiere, C.; Viallefont, P. *Tetrahedron* **1985**, *41*, 1833–1843.
- The thioester from alanine was prepared according to: Neises, B.; Steglich, W. Angew. Chem., Int. Ed. Engl. 1978, 17, 522–523.
- 10. For the Pd₂(dba)₃ use of higher reaction temperatures or more catalyst gave poorer results.
- The starting material was recovered, or when the reaction conditions were forced, the hydrolysis of carbamates led to polymeric mixtures.
- 12. Schmidt, U.; Meyer, R.; Leitenberger, V.; Stäbler, F.; Lieberknecht, A. *Synthesis* **1991**, 409–413.
- Gordon, S.; Costa, L.; Incerti, M.; Manta, E.; Saldaña, J.;
 Domínguez, L.; Mariezcurrena, R.; Suescun, L. *Il Farmaco* 1997, 52, 603–608.